

# Thermostable $\beta$ -Agarase



Catalog No.	Product	Size
317-07123	Thermostable $\beta$ -Agarase	30 units
311-07121		300 units

For research use only.

$\beta$ -agarase is an enzyme which hydrolyzes  $\beta$ -1,4 linkages in agarose to produce neoagaro-oligosaccharides. Agarose digested by  $\beta$ -agarase does not gelate again, therefore, nucleic acids can be recovered from agarose gels. Thermostable  $\beta$ -Agarase has a higher thermostability and stronger hydrolyzing activity than conventional  $\beta$ -agarase. Furthermore, a simple protocol allows for quick DNA and RNA purification. This enzyme is particularly suitable for purification of intact large DNA.

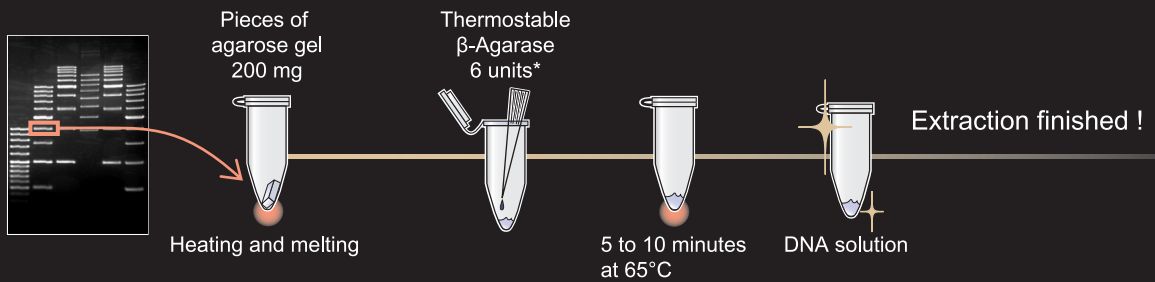
This enzyme was isolated from an agar-degrading bacterium living in a deep-sea by the Japan Agency for Marine-Earth Science and Technology (JAMSTEC) using the submersible SHINKAI6500.

- Activity: 1,000 units/ml
- Unit definition: 1 unit of Thermostable  $\beta$ -Agarase produces reducing sugar equivalent to 1  $\mu$ mol of D-galactose in 1 minutes at 60°C
- Form: 20 mM Tris-HCl (pH 7.5 at 25°C)  
50 mM NaCl
- Storage condition: Store at 2 to 8°C.

## Overview

- > Simple and short protocol (Reaction is completed in only 10 minutes).
- > Can be used on standard agarose as well as low melting point agarose.
- > Hydrolyzing gel solution is directly available for various applications. Such as cloning, restriction endonuclease digestion, sequencing, etc.
- > Can effectively extract intact large DNA from agarose gels.

## Procedure



\*The amount of an enzyme may be reduced by the density of the agarose gel.

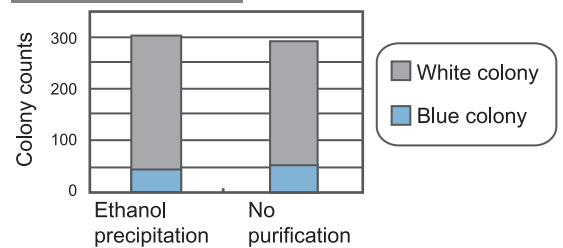
## Data

### Cloning of the DNA recovered from agarose gel pieces.

The 500 bp DNA fragment derived from  $\lambda$ DNA was amplified by PCR. PCR product were separated by 3% Agarose 21 (low molecular weight separation; NIPPONGENE).

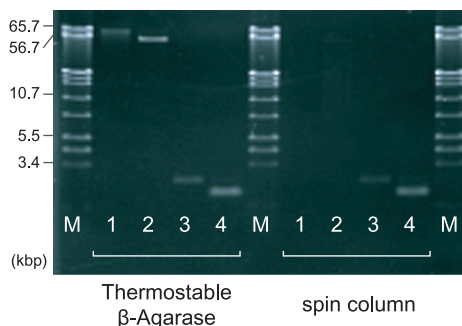
A required band was excised from the gel with a razor. After melting gel piece, it was treated by Thermostable  $\beta$ -Agarase. 9  $\mu$ l of the gel solution was added to 1  $\mu$ l of cloning vector DNA (3 kbp, 50 ng/ $\mu$ l) to set up ligation reaction using Ligation-Convenience Kit (NIPPON GENE). After ligation reaction, ECOS™ Competent *E.coli* DH5 $\alpha$  (NIPPON GENE) were transformed and then the number of colonies were counted. As the control, DNA solution purified by ethanol precipitation was used for ligation reaction.

### Cloning efficiency



### Extraction of intact large DNA from agarose gels.

T4 GT7 DNA (166 kbp),  $\lambda$ DNA (48.5 kbp), pUC19 DNA (2.69 kbp) separated by agarose gel electrophoresis in 1x TAE was excised from 0.3% Agarose L (low-melting point; NIPPON GENE). The gel pieces were treated by Thermostable  $\beta$ -Agarase or spin column and the collected amount of DNA was compared. One third amount of DNA was separated using 0.3% Agarose H (high gel strength; NIPPON GENE).



M: Marker 8 GT (0.4  $\mu$ g)  
 1: T4 GT7 DNA  
 2:  $\lambda$ DNA  
 3: pUC19 DNA (open circular)  
 4: pUC19 DNA (closed covalently circular)

# Questions and Answers



Q1. How amount of Thermostable  $\beta$ -Agarase should be used ?

A1. Generally, add the enzyme at a ratio of 6 unit (6  $\mu$ l) per 200 mg agarose gel. The amount of the enzyme can sometimes be reduced by the density of the gel and the condition.

- The amount of Thermostable  $\beta$ -Agarase which is needed for agarose gel to be hydrolyzed in 5 minutes

Agarose gel: 200mg

Agarose conc.	Enzyme volume
1.0% Agarose S	2.0 $\mu$ l (2.0 unit)
1.0% Agarose S	3.0 $\mu$ l (3.0 unit)
2.0% Agarose S	5.0 $\mu$ l (5.0 unit)
1.5% Agarose XP	3.0 $\mu$ l (3.0 unit)

- The amount of Thermostable  $\beta$ -Agarase which is needed for agarose gel to be hydrolyzed in 10 minutes

Agarose gel: 200 mg

Agarose conc.	Enzyme volume
1.5% Agarose S	1.5 $\mu$ l (1.5 unit)
2.0% Agarose S	3.0 $\mu$ l (3.0 unit)
3.0% Agarose 21	5.5 $\mu$ l (5.5 unit)

Related agarose products

Agarose	Melting point	Chracteristic
Agarose S	$\leq 90^{\circ}\text{C}$ (1.5%)	Standard agarose
Agarose HS	$\leq 93^{\circ}\text{C}$ (1.5%)	High gel strength type of Agarose S
Agarose 21	$\leq 85^{\circ}\text{C}$ (3.0%)	Low molecular weight separation
Agarose XP	$\leq 70^{\circ}\text{C}$ (3.0%)	Low-melting point and low molecular weight separation
Agarose X	$\leq 93^{\circ}\text{C}$ (4.0%)	High gel strength and low molecular weight separation
Agarose H	Boil (1.5%)	High gel strength and low molecular weight separation
Agarose L	$\leq 65^{\circ}\text{C}$ (1.5%)	Low-melting point
Agarose GB	$\leq 65^{\circ}\text{C}$ (1.5%)	For pulsed field gel electrophoresis

Q2. How can it be confirmed whether gel is completely hydrolyzed ?

A2. When not gelating even if DNA solution is cooled on the ice, agarose gel is completely hydrolyzed.

Q3. What is an advantage of thermostability ?

A3. It is not only used for standard gel, but can be used for low-melting point gel. Reaction finishes in only 10 minutes.

Q4. How much is recovery efficiency of a DNA ?

A4. Almost all of DNA can be recovered in general.

Q5. Thermostable  $\beta$ -Agarase maintains the enzyme activity at a long term ?

A5. This enzyme is stable for 2 years at 4 or  $-20^{\circ}\text{C}$ . When keeping in the room temperature, it's stable for 1 year. The activity is not altered during one-hundred cycles of freezing and thawing.

Q6. What amount of hydrolyzation solution produces ?

A6. When using 200 mg agarose gel, the amount of hydrolyzed solution will be 200  $\mu$ l.

Q7. How should hydrolyzed solution be concentrated?

A7. DNA can be concentrated by ethanol precipitation.

Q8. Can hydrolyzed solution be used for *in vitro* transcription reaction ?

A8. It's possible to synthesize RNA by using hydrolyzed solution directly as the template. CUGA<sup>®</sup>7 *in vitro* Transcription Kit (NIPPON GENE) was used for RNA synthesis.

Q9. Can hydrolyzed solution be used for DNA sequencing ?

A9. It's possible to sequence by using hydrolyzed solution directly as the template.

Q10. Is it possible to recover RNA from denaturing agarose gel by using Thermostable  $\beta$ -Agarase?

A10. It can be used, however formaldehyde inhibits the enzyme activity. Therefore, it's necessary to increase the amount of the enzyme.

## Information

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